



## Letter to the Editor

### Emergence of azithromycin resistance mediated by the *mph(A)* gene in *Salmonella* Typhimurium clinical isolates in Latin America



Sir,

*Salmonella enterica* represents one of the leading causes of food-borne diseases, constituting an important public-health problem worldwide. The global burden of disease caused by *Salmonella* infections has been estimated at more than 90 million human cases per year [1]. In Argentina between 2009–2016, *S. enterica* serotype Typhimurium was the most prevalent serotype, with a mean of 25%, followed by *S. enterica* serotype Enteritidis, with a mean of 14% (personal communication, National Reference Laboratory for *Salmonella*). Ceftriaxone and ciprofloxacin are the treatments of choice for invasive *Salmonella* infections, whilst azithromycin is suggested as an additional option [1]. The goal of this study was to characterise azithromycin-resistant *S. Typhimurium* clinical isolates harbouring the *mph(A)* gene isolated in Argentina.

A total of 12 *S. Typhimurium* isolates were recovered from faeces (10), psoas abscess (1) and blood (1) samples derived from six hospitals in Buenos Aires, Córdoba and Santa Fe Provinces (Table 1). Serovar determination was conducted according to the 9th edition of the White–Kauffmann–Le Minor scheme. Antimicrobial susceptibility testing was performed and was interpreted according to Clinical and Laboratory Standards Institute (CLSI) guidelines (M100–S25). All *S. Typhimurium* isolates showed similar multidrug-resistant (MDR) profiles, including high-level azithromycin resistance with MIC<sub>50/90</sub> values of 64/128 µg/mL (MIC range 32–128 µg/mL). Using the azithromycin breakpoints for *S. enterica* serotype Typhi defined for agar dilution by the CLSI in 2015 ( $\leq 16$  µg/mL, susceptible;  $\geq 32$  µg/mL, resistant), all 12 *S. Typhimurium* isolates were correctly categorised as resistant. All isolates were positive by standard PCR for the *mph(A)* gene. High azithromycin MICs ( $>16$  µg/mL) were previously reported in *S. enterica*, but only two *S. Typhimurium* isolates harbouring the *mph(A)* gene, recovered from the USA and the UK, have been described [2,3].

Seven strains were also resistant to cefotaxime and ceftazidime, which correlated with the presence of the *bla*<sub>CTX-M-14</sub> gene (Table 1). Extended-spectrum cephalosporin-resistant *Salmonella* clinical isolates have been recognised since the mid-1980 in isolates from Argentina [4], reaching 3.7% (15/402 isolates) in 2015, (WHONET–Argentina network; <http://antimicrobianos.com.ar/ATB/wp-content/uploads/2016/12/Informe-Resistencia-ARGENTINA-2015.pdf>). Intermediate resistance to ciprofloxacin was observed in 5 of the 12 isolates, which were positive for the *qnrB19* gene (Table 1). Four isolates showed

susceptibility only to fosfomycin, tigecycline, colistin and imipenem.

The 12 isolates were grouped by pulsed-field gel electrophoresis (PFGE) with the restriction enzyme *Xba*I into five clusters (I–V) and were compared with the PulseNet National Database (PNND), which include  $>1600$  *S. Typhimurium* patterns since 2004. Four isolates from hospital F from Córdoba Province were grouped as cluster III and were related to other isolates in the PNND recovered between 2012–2016 from Santa Fe and Buenos Aires Provinces. Cluster IV belonged to a pattern shared with another 224 isolates found in the PNND recovered from 10 provinces in the period 2009–2016, showing that this clone was already circulating across the country. Profiles of clusters I, II and V were not found in the database.

Whole-genome sequencing of the first MDR *S. Typhimurium* isolate (M17330) was performed using an Illumina compact MiSeq system (Illumina Inc., San Diego, CA). Using CLC Genomics Workbench software (QIAGEN), a total of 78 contigs were assembled. Contig\_33 (15 496 bp) revealed the presence of the *mph(A)*–*mrx*–*mphR(A)* cluster and showed 99% identity with a 16 258-bp fragment of p1 plasmid from *Klebsiella pneumoniae* KPN\_KPC\_HUG\_07 (NZ\_CP019773). This contig also contains a *catA1* gene and a class 1 integron with *dfrA12* and  $\Delta$ *aadA2* (763 bp deletion) cassettes. Others resistance genes, which correlated with its resistance phenotype, were detected in six contigs: *bla*<sub>CTX-M-14</sub>; *bla*<sub>TEM-1</sub>; *sul2*; *tetB*; *aph(3'')-Ib*; *aph(6)-Id*; *aph(3')-Ia*; *aac(6')-Iy*; and *qnrB19*.

Using sequence data and *S1* nuclease pulsed-field gel electrophoresis (*S1*-PFGE) analysis, four plasmids were identified in *S. Typhimurium* M17330: pM17330-1 (ca. 210 kb); pM17330-2 (ca. 106 kb); pM17330-3 (ca. 54 kb); and pM17330-4 (ca. 2.7 kb). Two *Escherichia coli* transconjugants (TCs) showing different susceptibility phenotypes were obtained by biparental conjugation (Table 1). *Escherichia coli* TC17330-1 was selected using sodium azide plus ampicillin, was resistant to cefotaxime and harboured the *bla*<sub>CTX-M-14</sub> gene in the pM17330-2 plasmid. As both *catA1* and *mph(A)* genes were present in the same assembled contig, *E. coli* TC17330-2 was selected using sodium azide plus chloramphenicol. TC17330-2 strain harboured pM17330-1 and pM17330-2 plasmids and showed resistance to azithromycin, ampicillin, cefotaxime, trimethoprim/sulfamethoxazole, kanamycin, streptomycin and tetracycline. Contig\_24, corresponding to pM17330-4, shared 100% similarity with pPAB19-4 plasmid harbouring the *qnrB19* gene and previously described in *Salmonella* sp. M9397 isolate from Argentina (JN995611).

*Salmonella* Typhimurium represents the most frequent serotype in Argentina, and the presence of the *mph(A)* gene in isolates from different locations and different clones, that are located on conjugative plasmids harbouring additional resistance genes is worrisome for the health system and requires continuous surveillance of azithromycin susceptibility in this *Salmonella*

**Table 1**  
Epidemiological data, susceptibility profiles and antimicrobial resistance genes of 12 *Salmonella enterica* serotype Typhimurium isolates.

Isolate number	Hospital	Province	Isolation date	Specimen	MIC ( $\mu\text{g}/\text{mL}$ )							Acquired resistance genes	Xbal-PFGE profile
					AZM	AMP	CAZ	CTX	IPM	NAL	CIP		
M17330	A	Santa Fe	04/11/2014	Psoas abscess	64	$\geq 256$	4	128	0.25	32	0.5	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub> , <i>qnrB19</i>	II
M17353	B	Buenos Aires	05/07/2014	Stool	64	$\geq 256$	4	128	0.25	32	0.5	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub> , <i>qnrB19</i>	V
M17528	D	Santa Fe	06/12/2014	Stool	128	$\geq 256$	4	64	0.25	32	0.5	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub> , <i>qnrB19</i>	IV
M17891	E	Santa Fe	07/21/2014	Stool	64	$\geq 256$	4	128	0.12	4	0.03	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub>	IV
M17671	D	Santa Fe	08/16/2014	Stool	32	$\geq 256$	4	128	0.25	4	0.015	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub>	I
M17728	D	Santa Fe	08/30/2014	Stool	64	$\geq 256$	4	128	0.25	4	0.03	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub>	V
M17767	F	Córdoba	09/09/2014	Stool	64	$\geq 256$	0.25	0.06	0.25	4	0.03	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub>	III
M17768	F	Córdoba	09/10/2014	Stool	64	$\geq 256$	0.25	0.12	0.25	4	0.03	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub>	III
M17769	F	Córdoba	09/11/2014	Stool	64	$\geq 256$	0.25	0.12	0.25	4	0.03	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub>	III
M17770	F	Córdoba	09/13/2014	Stool	64	$\geq 256$	0.5	0.06	0.25	4	0.03	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub>	III
M17945	F	Córdoba	12/07/2014	Blood	64	$\geq 256$	0.25	0.06	0.12	32	0.5	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>qnrB19</i>	I
M17967	C	Buenos Aires	12/07/2014	Stool	128	$\geq 256$	16	$\geq 256$	0.25	32	0.5	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub> , <i>qnrB19</i>	V
TC17330-1	N/A	N/A	N/A	N/A	2	$\geq 256$	0.25	16	0.12	4	0.03	<i>bla</i> <sub>CTX-M-14</sub>	N/A
TC17330-2	N/A	N/A	N/A	N/A	32	$\geq 256$	0.12	16	0.12	4	0.015	<i>mph(A)</i> , <i>bla</i> <sub>TEM-1</sub> , <i>bla</i> <sub>CTX-M-14</sub>	N/A
<i>Escherichia coli</i> J53	N/A	N/A	N/A	N/A	2	4	0.12	0.06	0.12	4	0.03	–	N/A

MIC, minimum inhibitory concentration; AZM, azithromycin; AMP, ampicillin; CAZ, ceftazidime; CTX, cefotaxime; IPM, imipenem; NAL, nalidixic acid; CIP, ciprofloxacin; N/A, not applicable; PFGE, pulsed-field gel electrophoresis.

All *S. Typhimurium* clinical isolates plus the transconjugant *E. coli* TC17330-1 were also resistant to trimethoprim/sulfamethoxazole, chloramphenicol, tetracycline, kanamycin and streptomycin and were susceptible to fosfomicin, tigecycline, colistin and imipenem.

serotype. Since 2015, azithromycin was included to be routinely tested by the National Antimicrobial Surveillance–WHONET–Argentina network. Up to June 2017, 19 additional *S. Typhimurium* clinical isolates were identified as positive for the *mph(A)* gene. To the best of our knowledge, these represent the first azithromycin-resistant *S. Typhimurium* clinical isolates associated with the *mph(A)* gene in the Latin American region. Emergence of the *mph(A)* gene and its capacity for horizontal dissemination jeopardises the use of azithromycin for severe *Salmonella* infections.

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AZM-R-Group: Dra María Rosa Baroni (Hospital de Niños ‘Dr O. Allasia’, Santa Fe); Dra Noemi Borda (Hospital Español de Rosario, Santa Fe); Dra María Cristina Guardati (Hospital Celemene Alvarez, Santa Fe); Dra Liliana Gonzalez (Hospital Infantil Municipal de Córdoba, Córdoba); Dra Viviana Vilchez (Hospital Universitario Austral, Buenos Aires); and Dra Miryam Vazquez (Hospital de Niños ‘Dr R. Gutierrez’, Ciudad Autónoma de Buenos Aires).

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#### Competing interests

None declared.

#### Ethical approval

Not required.

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Diego Faccone<sup>a,b</sup>

<sup>a</sup>Servicio Antimicrobianos, National Reference Laboratory in Antimicrobial Resistance (NRLAR), Instituto Nacional de Enfermedades Infecciosas (INEI)–ANLIS ‘Dr C. Malbrán’, Buenos Aires, Argentina

<sup>b</sup>Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Buenos Aires, Argentina

Celeste Lucero<sup>a</sup>

Ezequiel Albornoz<sup>a</sup>

Alejandro Petroni<sup>a</sup>

Paola Ceriana<sup>a</sup>

<sup>a</sup>Servicio Antimicrobianos, National Reference Laboratory in Antimicrobial Resistance (NRLAR), Instituto Nacional de Enfermedades Infecciosas (INEI)–ANLIS ‘Dr C. Malbrán’, Buenos Aires, Argentina

Josefina Campos<sup>c</sup>

María Rosa Viñas<sup>c</sup>

<sup>c</sup>Servicio Enterobacterias, National Reference Laboratory for *Salmonella* (NRL for *Salmonella*), INEI–ANLIS ‘Dr C. Malbrán’, Buenos Aires, Argentina

Geraldine Francis<sup>a</sup>AZM-R-Group

<sup>a</sup>Servicio Antimicrobianos, National Reference Laboratory in Antimicrobial Resistance (NRLAR), Instituto Nacional de Enfermedades Infecciosas (INEI)–ANLIS ‘Dr C. Malbrán’, Buenos Aires, Argentina

Roberto G. Melano<sup>d</sup>

<sup>d</sup>Public Health Ontario Laboratories, Toronto, ON, Canada

Alejandra Corso<sup>a,\*</sup>

<sup>a</sup>Servicio Antimicrobianos, National Reference Laboratory in Antimicrobial Resistance (NRLAR), Instituto Nacional de Enfermedades Infecciosas (INEI)–ANLIS ‘Dr C. Malbrán’, Buenos Aires, Argentina

\* Corresponding author.

E-mail address: [acorso@anlis.gov.ar](mailto:acorso@anlis.gov.ar) (A. Corso).

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